

# A DNA barcode library for ground beetles of Germany: the genus *Agonum* Bonelli, 1810 (Insecta, Coleoptera, Carabidae)

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## Abstract

The ground beetle genus *Agonum* Bonelli, 1810 is a large genus of the tribe Platynini with many species that show high amounts of intraspecific variations, making a correct identification challenging. As part of the German Barcode of Life initiative, this publication provides a comprehensive DNA barcode library for species of *Agonum* that are reported for Germany. In total, DNA barcodes from 258 beetles and 23 species were analysed using the Barcode of Life Data System (BOLD) workbench, including sequences from former studies and 68 newly-generated sequences. The neighbour-joining analyses, based on K2P distances, revealed distinct clustering for all studied species, with unique Barcode Index Numbers (BINs) for 15 species (65%). BIN sharing but distinct clustering was found for three species pairs: *Agonum micans*/*Agonum scitulum*, *Agonum impressum*/*Agonum sexpunctatum* and *Agonum duftschmidi*/*Agonum emarginatum*. The given dataset and its analysis represent another important step in generating a comprehensive DNA barcode library for the ground beetles of Germany and Central Europe in terms of modern biodiversity research.

## Key Words

BOLD, Central Europe, cytochrome *c* oxidase subunit I, German Barcode of Life, mitochondrial DNA, molecular specimen identification, sibling species

## Introduction

Species identification represents a pivotal component for biodiversity studies and conservation planning, but represents a challenge for many taxa when using morphological traits only (e.g. the correct identification of juveniles or larval stages). As a consequence of tremendous technological advances in molecular biology during the last 20 years, molecular data have become increasingly popular in species identification. In this context, DNA barcoding represents the central component in the modern diagnostic toolbox of molecular biodiversity assessment studies and taxonomic research (e.g. Hebert and Gregory 2005; Krees et al. 2015; Hajibabaei et al. 2016; Miller et al. 2016). For animals, a 658 base pair (bp) fragment of the mitochondrial

cytochrome *c* oxidase subunit I (COI) gene has been selected as a DNA barcode (Hebert et al. 2003a, b). The utility of DNA barcoding relies on the assumption that genetic variation within a species is much smaller than variation between species. Barcode sequences are typically deposited in the international Barcode of Life Data Systems database (BOLD; <http://www.boldsystems.org>). This public database acts as the central core data interface and repository that allows researchers to collect, to organise and to analyse DNA barcode data (Ratnasingham and Hebert 2007). In addition to various analytical tools implemented in the BOLD workbench, DNA barcodes can be analysed using the Barcode Index Number (BIN) system that clusters DNA barcodes to produce operational taxonomic units that closely correspond to species (Ratnasingham and Hebert 2013).



Nevertheless, it should be noted that various effects can seriously limit the use of DNA barcoding or mitochondrial markers in general, in terms of molecular species identification (e.g. Will and Rubinoff 2004; Will et al. 2005; Rubinoff et al. 2006; Collins and Cruickshank 2013; Duran et al. 2019). For example, closely related but distinct species, as well as species that hybridise, may share identical haplotypes (e.g. Sota and Vogler 2002; Takami and Suzuki 2005; Andujar et al. 2014). In contrast to this, complex phylogeographic histories (e.g. Schoville et al. 2012; Faille et al. 2015; Weng et al. 2016) and incomplete lineage sorting effects (e.g. Sota et al. 2005; Zhang et al. 2005; Weng et al. 2020) can contort the mitochondrial variability of the studied organisms, generating para- and polyphyly in phylogenetic trees (Funk and Omland 2003; Ross 2014; but see Mutanen et al. 2016). Heteroplasmy (Boyce et al. 1989) or the presence of mitochondrial pseudogenes (Hazakani-Covo et al. 2010; Maddison 2012) can affect a successful amplification of the target fragment. Finally, maternally-inherited endosymbionts, such as the alpha-proteobacteria *Wolbachia*, may cause a linkage disequilibrium within mtDNA in arthropods and result in a homogenisation of mtDNA haplotypes (e.g. Hurst and Jiggins 2005; Kolasa et al. 2018; Kajtoch et al. 2019). However, a vast number of studies across a broad range of taxa demonstrate the efficiency of DNA barcoding as the method of choice in molecular species identification (e.g. Raupach et al. 2014; Schmidt et al. 2015; Morinière et al. 2017; Schmid-Egger et al. 2019).

Not surprisingly, the build-up of comprehensive DNA barcode libraries represents a pivotal task (e.g. Brandon-Mong et al. 2015; Curry et al. 2018; Weigand et al. 2019). In Germany, the German Barcode of Life initiative (GBOL; [www.bolgermany.de](http://www.bolgermany.de)) aims to assess the genetic diversity of all animals, fungi and plants of Germany. Numerous studies provided first comprehensive DNA barcode libraries for various insect taxa (e.g. Hausmann et al. 2011; Hendrich et al. 2015; Hawlitschek et al. 2017; Havemann et al. 2018). In the case of ground beetles (Carabidae), a number of previous studies started to build up a comprehensive DNA barcode library for these routinely-used biological indicators, including the genera *Bembidion* Latreille, 1802 (Raupach et al. 2011; Raupach et al. 2016), *Amara* Bonelli, 1810 (Raupach et al. 2018) and *Notiophilus* Duméril, 1806 (Raupach et al. 2019). Detailed studies of many other important genera, however, are still missing.

Within the Carabidae, the genus *Agonum* Bonelli, 1810 is a species-rich taxon of shiny black or metallic ground beetles that imitate large representatives of *Bembidion* Latreille, 1802 (Lindroth 1986) (Fig. 1). Beetles of this genus are commonly found in forested and open habitats adjacent to freshwater throughout Europe, the eastern Palearctic area and North America (Liebherr and Schmidt 2004; Bousquet 2012). They are generalist predators that feed on a wide range of small arthropods, for example, springtails, aphids or midges (e.g. Griffiths et al. 1985; Bilde and Toft 1994; Hannam et al. 2008).

A first detailed phylogenetic analysis of all species of the genus *Agonum* that was based on numerous morphological characters recognised four subgeneric entities: *Agonum* sensu stricto, *Europhilus* Chaudoir, 1859, *Olisares* Motschulsky, 1865 and *Platynomicrus* Casey, 1920 (Liebherr and Schmidt 2004; Liebherr et al. 2005). Whereas some species are distinctive, others are difficult to identify as a consequence of intraspecific variations in setation, pronotal shape, body colouration and size and, therefore, typically require the examination of the genitalia (Hürka 1996; Luff 2007). In the case of the *Agonum* species that are documented for Germany, the combination of various characteristic traits, including, besides others, a well-developed median tooth at the mentum and the shape of the posterior angles of the pronotum, allows a correct identification (Schmidt, pers. communication). In Europe, species related to *Agonum viduum* are most challenging in terms of identification, but this group has been carefully revised more than 25 years ago (Schmidt 1994). For Germany, 24 species have been recorded so far (Trautner et al. 2014; Schmidt et al. 2016). As a consequence of many *Agonum* species being found in highly-endangered peat bogs or similar freshwater-associated habitats, numerous species have become very rare or even threatened with extinction, for example *Agonum hypocrita* (Apfelbeck, 1904) or *Agonum munsteri* (Hellén, 1935) (Schmidt et al. 2016).

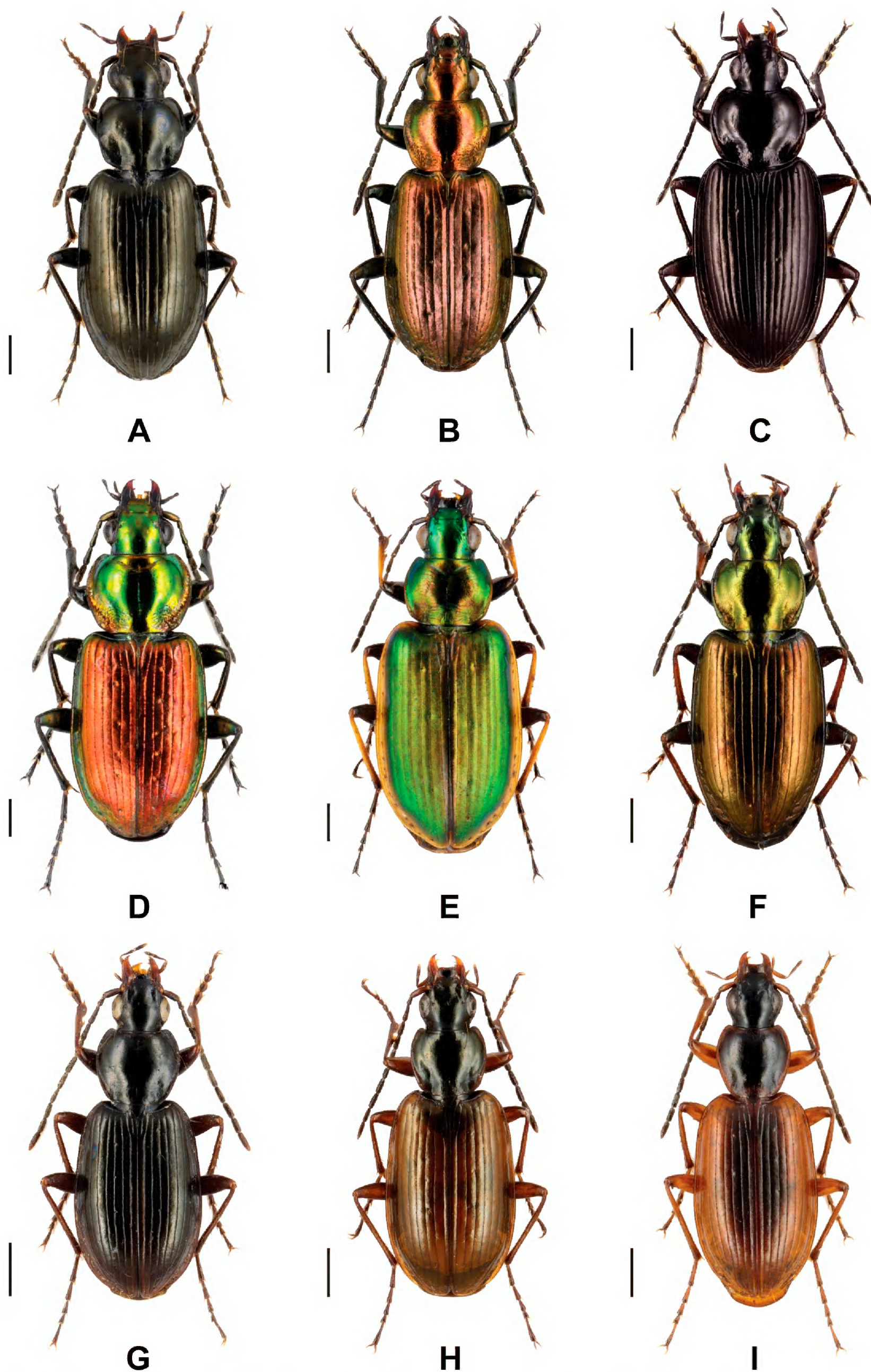
In this study, we present, as part of the on-going GBOL project, another step in generating a comprehensive DNA barcode library for the molecular identification of Central European ground beetle species, here focusing on the genus *Agonum*. The analysed sequence library included 23 species of *Agonum* and *Oxypselaphus obscurus* (Paykull, 1790) as outgroup. We generated 68 new barcodes including some sequences of old pinned museum specimens and analysed a total number of 258 DNA barcodes in detail.

## Material and methods

### Sampling of specimens

Most of the sampled ground beetles ( $n = 186$ , 78%) were collected between 1999 and 2015 using various sampling methods (i.e. hand collecting, pitfall traps). All beetles were stored in ethanol (96%). The analysed specimens were identified using the identification key provided in Schmidt (2006). It was also possible to generate DNA barcodes from decade-old pinned ground beetles of the carabid collection of the Bavarian State Collection of Zoology, namely *Agonum antennarium* (Duftschmid, 1812) ( $n = 2$ ) with an age of 87 years and *A. impressum* (Panzer, 1796) ( $n = 1$ , age: 52 years). In total, 68 new barcodes of 17 species were generated, including five species new to BOLD. For our analysis, we also included 175 DNA barcodes of four previous studies (Raupach et al. 2010: 17 specimens, 5 species; Hendrich et al. 2015: 101 specimens, 13 species; Pentinsaari et al. 2014: 52 specimens, 12 species;





**Figure 1.** Representative images of analysed *Agonum* species. **A.** *Agonum (Olisares) duftschmiedi* Schmidt, 1994; **B.** *Agonum (Olisares) ericeti* (Panzer, 1809); **C.** *Agonum (Olisares) hypocrita* (Apfelbeck, 1904); **D.** *Agonum (Olisares) sexpunctatum* (Linné, 1758); **E.** *Agonum (Agonum) marginatum* (Linné, 1758); **F.** *Agonum (Agonum) muelleri* (Herbst, 1784); **G.** *Agonum (Europhilus) fuliginosum* (Panzer, 1809); **H.** *Agonum (Europhilus) piceum* (Linné, 1758) and **I.** *Agonum (Europhilus) thoreyi* Dejean, 1828. Scale bars: 1 mm. All images were obtained from <http://www.eurocarabidae.de>.



Rulik et al. 2018: 5 specimens, 1 species), as well as 15 public barcodes deposited in BOLD without corresponding publication (10 species).

Most of the studied beetles were collected in Germany ( $n = 191$ , 73.9%), but for comparison, a number of specimens were included from Austria ( $n = 10$ , 3.6%), Czech Republic ( $n = 1$ , 0.4%), Estonia ( $n = 1$ , 0.4%), Finland ( $n = 49$ , 18.9%), France ( $n = 2$ , 0.8%), Italy ( $n = 1$ , 0.4%), Montenegro ( $n = 2$ , 0.8%) and Sweden ( $n = 2$ , 0.8%). The number of analysed specimens per species ranged from one (*A. gracilipes* (Duftschmid, 1812), *A. impressum* (Panzer, 1796)) to a maximum of 31 for *A. fuliginosum* (Panzer, 1809).

## DNA barcode amplification, sequencing and data depository

All laboratory operations were carried out, following standardised protocols for COI amplification and sequencing (Ivanova et al. 2006; deWaard et al. 2008), at the Canadian Center for DNA Barcoding (CCDB), University of Guelph, the molecular labs of the Zoologisches Forschungsmuseum Alexander Koenig (ZFMK) in Bonn and the working group Systematics and Evolutionary Biology at the Carl von Ossietzky University Oldenburg, the latter two being located in Germany. Photos from each studied beetle were taken before molecular work started. One or two legs of one body side were removed for the subsequent DNA extraction which was performed using the QIAmp Tissue Kit (Qiagen GmbH, Hilden, Germany) or NucleoSpin Tissue Kit (Macherey-Nagel, Düren, Germany), following the extraction protocol.

Detailed information about used primers, PCR amplification and sequencing protocols can be found in a previous publication (see Raupach et al. 2016). All purified PCR products were cycle-sequenced and sequenced in both directions by contract sequencing facilities (Macrogen, Seoul, South Korea or GATC, Konstanz, Germany), using the same primers as used in PCR. Double-stranded sequences were assembled and checked for mitochondrial pseudogenes (numts) by analysing the presence of stop codons and frameshifts, as well as double peaks in chromatograms with the Geneious Prime 2020.0.4 (<https://www.geneious.com>) (Biomatters, Auckland, New Zealand). For verification, BLAST searches (nBLAST, search set: others, programme selection: megablast) were conducted to confirm the identity of all new sequences based on already published sequences (high identity values, very low E-values).

Detailed voucher information, taxonomic classifications, photos, DNA barcode sequences, primer pairs used and trace files (including their quality) were uploaded to the public dataset “DS-BAAGO” (Dataset ID: [dx.doi.org/10.5883/DS-BAAGO](https://dx.doi.org/10.5883/DS-BAAGO)) on the Barcode of Life Data Systems (BOLD; [www.boldsystems.org](http://www.boldsystems.org)) (Ratnasingham and Hebert 2007). Parallel to this, all new barcodes were deposited in GenBank (accession numbers: MT520822–MT520889).

## DNA Barcode analysis

The complete dataset was analysed by using various approaches. First, the comprehensive analysis tools of the BOLD workbench were employed to calculate the nucleotide composition of the sequences and distributions of Kimura-2-parameter distances (K2P; Kimura 1980) within and between species (align sequences: BOLD aligner; ambiguous base/gap handling: pairwise deletion). In addition, all barcode sequences were subjected to the Barcode Index Number (BIN) analysis system implemented in BOLD that clusters DNA barcodes in order to produce operational taxonomic units that typically closely correspond to species (Ratnasingham and Hebert 2013). A threshold of 2.2% was applied for a rough differentiation between intraspecific and interspecific distances, based on Ratnasingham and Hebert (2013). These BIN assignments on BOLD are constantly updated as new sequences are added, splitting and/or merging individual BINs in the light of new data.

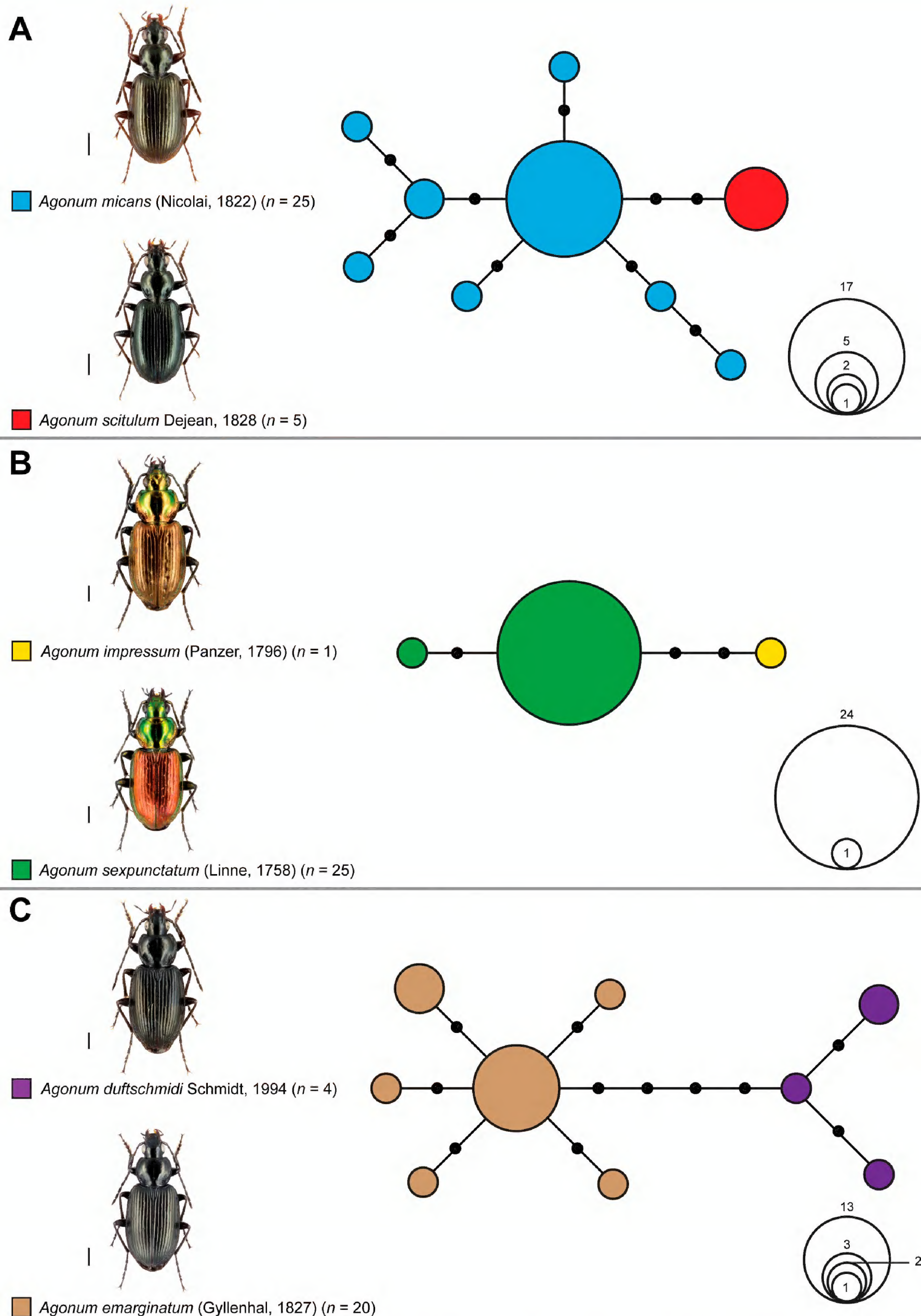
Second, maximum parsimony networks were constructed with TCS 1.21, based on default settings (Clement et al. 2000) and implemented in the software package PopART v.1.7 (Leigh and Bryant 2015), in the case of species pairs with interspecific distances smaller than 2.2% and sharing identical BINs. Such networks allow a better visualisation of the distances between closely-related species than classical tree topologies.

Finally, all sequences were aligned using MUSCLE (Edgar 2004) and analysed using a neighbour-joining cluster analysis (NJ; Saitou and Nei 1987), based on K2P distances with MEGA 10.0.5 (Kumar et al. 2018). Non-parametric bootstrap support values were obtained by re-sampling and analysing 1,000 pseudo-replicates (Felsenstein 1985). It should be noted that DNA barcodes do not aim to recover phylogenetic relationships (e.g. DeSalle and Goldstein 2019). Instead, the shown topology represents a graphical visualisation of DNA barcode distance divergences and putative species cluster.

## Results

Overall, 252 DNA barcode sequences of 24 *Agonum* species were analysed, representing 96% of all documented species ( $n = 25$ ) of this genus for Germany, except *Agonum nigrum* Dejean, 1828. A full list of the analysed species is presented in the supporting information (Suppl. material 1). Lengths of the analysed DNA barcode fragments ranged from 307 to 658 base pairs (bp). As is typically known for arthropods, the DNA barcode region has a high AT-content (68%), with mean sequence compositions for Adenosine (A) = 31%, Cytosine (C) = 16%, Guanine (G) = 16% and Tyrosine (T) = 37%. Intraspecific K2P distances ranged from 0 to a maximum of 1.3% (*Agonum ericeti* (Panzer, 1809)), whereas interspecific distances had values between 0.16 and 7.07%. Lowest interspecific distances were found for *Agonum micans*





**Figure 2.** Maximum statistical parsimony networks of the species pairs. **A.** *Agonum micans* (Nicolai, 1822) (blue) and *Agonum scitulum* Dejean, 1828 (red); **B.** *Agonum impressum* (Panzer, 1796) (yellow) and *Agonum sexpunctatum* (Linne, 1758) (green) and **C.** *Agonum duftschmidi* Schmidt, 1994 (violet) and *Agonum emarginatum* (Gyllenhal, 1827) (light brown). Used parameters included default settings for connection steps, gaps were treated as fifth state. Each line represents a single mutational change, whereas small black dots indicate missing haplotypes. The numbers of analysed specimens ( $n$ ) are listed, whereas the diameter of the circles is proportional to the number of haplotypes sampled (see given open circles with numbers). Scale bars: 1 mm. Beetle images were obtained from <http://www.eurocarabidae.de>.



(Nicolai, 1822) and *Agonum scitulum* Dejean, 1828) with a value of 0.16% and the same BIN (AAN9978).

In total, unique BINs were revealed for 15 species (65%) and one BIN for two species for three species pairs (13%), namely *Agonum micans* (Nicolai, 1822)/*Agonum scitulum* Dejean, 1828), *Agonum impressum* (Panzer, 1796)/*Agonum sexpunctatum* (Linne, 1758) and *Agonum duftschmidi* Schmidt, 1994/*Agonum emarginatum* (Gyllenhal, 1827) (Fig. 2). Due to the fact that the numbers of unspecified nucleotides (“Ns”) exceeds more than 1% of their total length (*Agonum antennarium* (Duftschmid, 1812),  $n = 2$ ) or only sequences < 500 bp were given (*Agonum ericeti* (Panzer, 1809),  $n = 2$ ), two species received no BIN assignment.

The NJ analyses, based on K2P distances, revealed non-overlapping clusters for all analysed species (Fig. 3). In the case of species with more than two analysed specimens ( $n = 22$ ), 82% ( $n = 18$ ) of the studied species were characterised with bootstrap support values > 95%. A more detailed topology of all analysed specimens is presented in the supporting information (Suppl. material 2).

## Discussion

The results of this study highlight the efficiency of DNA barcodes for the determination of most German species of the genus *Agonum*, but also indicate a close relationship of a number of species with shared BINs, for example, *Agonum scitulum* (Dejean, 1828) and *Agonum micans* (Nicolai, 1822) (Suppl. material 1, Fig. 2). Haplotypes of both species are separated by a K2P distance of 0.16 and only three mutational steps (Fig. 2A). Despite the fact that *Agonum scitulum* has been often overlooked and misidentified as *Agonum micans* as a result of flawed identification keys in the past (e.g. Paill 2010; Schmidt and Benedikt 2010; Brigić et al. 2016), such a close relationship between these two species had, however, not been expected before (see Liebherr and Schmidt 2004). *Agonum scitulum* (Dejean, 1828) is a macropterous Western Palaearctic species with a discontinuous distribution from England to Romania, Croatia and the European part of Russia (Luff 2007; Paill 2010; Schmidt and Benedikt 2010; Brigić et al. 2016). It is a very rare species that is typically found in marshes, fens and reed beds, syntopic with *Agonum micans* and *Agonum fuliginosum* (Panzer, 1809) (Luff 2007; Paill 2010; Schmidt and Benedikt 2010). Thanks to a thorough review some years ago, an excellent key for the genus *Agonum* has been established and allows a reliable identification of both species, i.e. the absence/presence of hairs on the dorsal site of the 3<sup>rd</sup> tarsomere of the last walking leg (Schmidt 2006). Low molecular distances between both species give evidence for an apparent recent separation, but the small number of studied specimens, as well as the only use of mitochondrial sequence data, does not allow more conclusions at the moment. Here, additional analysis combining a careful morphological analysis, as well as detailed fine-scaling nuclear sequence data, will provide more information.

In contrast to the previous species pair, a close relationship between *Agonum impressum* (Panzer, 1796) and *Agonum sexpunctatum* (Linne, 1758), as well as *Agonum duftschmidi* Schmidt, 1994 and *Agonum emarginatum* (Gyllenhal, 1827), has been already suggested in the past (Liebherr and Schmidt 2004). The given barcode data support this hypothesis. Similar to the *Agonum micans* and *Agonum scitulum*, haplotypes of *Agonum impressum* (Panzer, 1796) and *Agonum sexpunctatum* (Linne, 1758) are separated by three mutational steps (K2P distance: 0.36) (Fig. 2B), whereas the minimum K2P distance of *Agonum duftschmidi* Schmidt, 1994 and *Agonum emarginatum* (Gyllenhal, 1827) has a value of 0.77, resulting in five additional mutational steps (Fig. 2C). Only the analysis of additional specimens from different locations will show if these distances within both species pairs are sound.

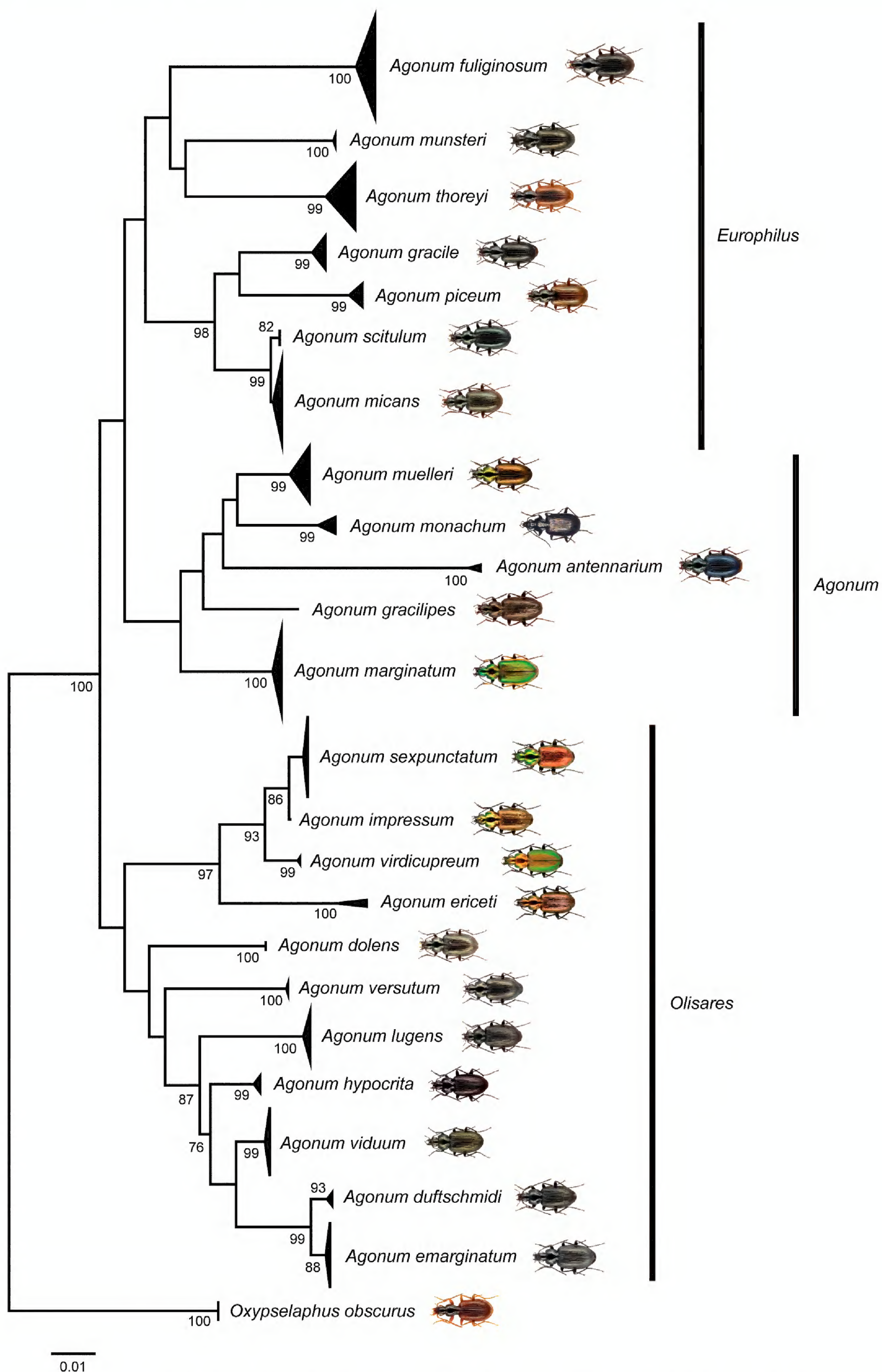
In comparison with other Central European ground beetles, for instance species of the genera *Bembidion* Latreille, 1802 (Raupach et al. 2016), *Calathus* Bonelli, 1810 (Hendrich et al. 2015) or *Notiophilus* Duméril, 1806 (Raupach et al. 2019), no interspecific distances > 2.2% or multiple BINs per species were found for the studied species and specimens. Highest intraspecific distances were identified for *Agonum ericeti* (Panzer, 1809) with a value of 1.3, *Agonum muelleri* (Herbst, 1784) (1.24) and *Agonum thoreyi* Dejean, 1828 (1.03) (Suppl. material 1). Nevertheless, all three species showed no conspicuous substructure for the analysed COI sequences yet.

At this point it should be kept in mind that not all beetles were collected in Germany. For instance, all specimens of *Agonum antennarium* (Duftschmid, 1812) were sampled in Montenegro (see Suppl. material 2), whereas beetles of other species were sampled from different localities in Germany only (e.g. *Agonum lugens* (Duftschmid, 1812) or *Agonum micans* (Nicolai, 1812)). As previously mentioned, a number of species are very rare in the target area, but more abundant in adjacent countries. In most cases, the new DNA barcodes represent, however, the very first molecular data for these taxa and give important impressions about their molecular diversity, even if they were not sampled in Germany. Nevertheless, it is also planned to analyse specimens for such species that were collected in Germany in the near future.

## Conclusion

As central part of modern biodiversity research, DNA barcoding will become more and more prominent in this research field. Therefore, comprehensive DNA barcode libraries of important bio-indicators will become the backbone of any applied analysis, for example, metabarcoding as well as eDNA studies. The present study demonstrates the successful identification of most *Agonum* species documented for Germany by using DNA barcodes. The dataset also shows a close relationship between various species and their putative recent origin. Only the analysis of additional species and specimens, however, will reveal if the observed patterns of distinct clusters persist.





**Figure 3.** Neighbour-joining (NJ) topology of the analysed ground beetle species, based on Kimura 2-parameter distances. Triangles show the relative number of individual's sampled (height) and sequence divergence (width). Numbers next to nodes represent non-parametric bootstrap values > 75% (1,000 replicates). Beetle images were obtained from <http://www.eurocarabidae.de> except for *Agonum monachum* (Duftschmid, 1812) (Photo taken by Lars Hendrich).



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## References

- Andujar C, Arribas P, Ruiz C, Serrano J, Gomez-Zurita J (2014) Integration of conflict into integrative taxonomy: fitting hybridization in species delimitation of *Mesocarabus* (Coleoptera: Carabidae). *Molecular Ecology* 23: 4344–4361. <https://doi.org/10.1111/mec.12793>
- Bilde T, Toft S (1994) Prey preference and egg production of the carabid beetle *Agonum dorsale*. *Entomologia Experimentalis et Applicata* 73: 151–156. <https://doi.org/10.1111/j.1570-7458.1994.tb01850.x>
- Bousquet Y (2012) Catalogue of Geadephaga (Coleoptera, Adephaga) of America, north of Mexico. *ZooKeys* 245: 1–1722. <https://doi.org/10.3897/zookeys.245.3416>
- Boyce TM, Zwick ME, Aquardo CF (1989) Mitochondrial DNA in the bark weevils: size, structure and heteroplasmy. *Genetics* 123: 825–836.
- Brandon-Mong GJ, Gan HM, Sing KW, Lee PS, Lim PE, Wilson JJ (2015) DNA metabarcoding of insects and allies: an evaluation of primers and pipelines. *Bulletin of Entomological Research* 105: 717–727. <https://doi.org/10.1017/S0007485315000681>
- Brigić A, Šoštarić R, Sedlar Z, Vučić-Karlo S (2016) *Agonum scitulum* Dejean, 1828 (Coleoptera, Carabidae) – new data on a rare carabid beetle species in Croatia. *Natura Croatia* 25: 279–285. <https://doi.org/10.20302/NC.2016.25.23>
- Clement M, Posada D, Crandall KA (2000) TCS: a computer program to estimate gene genealogies. *Molecular Ecology* 9: 1657–1660. <https://doi.org/10.1046/j.1365-294x.2000.01020.x>
- Collins RA, Cruickshank RH (2013) The seven deadly sins of DNA barcoding. *Molecular Ecology Resources* 13: 969–975. <https://doi.org/10.1111/1755-0998.12046>
- Curry CJ, Gibson JF, Shokralla S, Hajibabaei M, Baird DJ (2018) Identifying North American freshwater invertebrates using DNA barcodes: are existing COI sequence libraries fit for purpose? *Freshwater Science* 37: 178–189. <https://doi.org/10.1086/696613>
- DeSalle R, Goldstein P (2019) Review and interpretation of trends in DNA barcoding. *Frontiers in Ecology and Evolution* 7: e302. <https://doi.org/10.3389/fevo.2019.00302>
- deWaard JR, Ivanova NV, Hajibabaei M, Hebert PDN (2008) Assembling DNA barcodes: analytical protocols. In: Martin C (Ed.) *Methods in Molecular Biology: Environmental Genetics*. Humana Press, Totowa, 275–293. [https://doi.org/10.1007/978-1-59745-548-0\\_15](https://doi.org/10.1007/978-1-59745-548-0_15)
- Duran DP, Laroche RA, Gough HM, Gwiazdowski RA, Knisley CB, Herrmann DP, Roman SJ, Egan SP (2019) Geographic life history differences predict genomic divergence better than mitochondrial barcodes of phenotypes. *Genes* 11(3): e265. <https://doi.org/10.3390/genes11030265>
- Edgar RC (2004) MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics* 5: e113. <https://doi.org/10.1186/1471-2105-5-113>
- Faille A, Tänzler R, Toussaint EFA (2015) On the way to speciation: Shedding light on the Karstic phylogeography of the microendemic cave beetle *Aphaenops cerberus* in the Pyrenees. *Journal of Heredity* 106: 692–699. <https://doi.org/10.1093/jhered/esv078>
- Felsenstein J (1985) Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39: 783–791. <https://doi.org/10.1111/j.1558-5646.1985.tb00420.x>
- Funk DJ, Omland KE (2003) Species-level paraphyly and polyphyly: Frequency, causes, and consequences, with insights from animal mitochondrial DNA. *Annual Review of Ecology, Evolution, and Systematics* 34: 397–423. <https://doi.org/10.1146/annurev.ecolsys.34.011802.132421>
- Griffiths E, Wratten SD, Vickerman GP (1985) Foraging by the carabid *Agonum dorsale* in the field. *Ecological Entomology* 10: 181–189. <https://doi.org/10.1111/j.1365-2311.1985.tb00547.x>
- Hajibabaei M, Baird DJ, Fahner NA, Beiko R, Golding GB (2016) A new way to contemplate Darwin's tangled bank: how DNA barcodes are reconnecting biodiversity science and biomonitoring. *Philosophical Transactions of the Royal Society B: Biological Sciences* 371(1702): e20150330. <https://doi.org/10.1098/rstb.2015.0330>
- Hannam JJ, Lieberr JK, Hajek AE (2008) Climbing behaviour and aphid predation by *Agonum muelleri* (Coleoptera: Carabidae). *The Canadian Entomologist* 140: 203–207. <https://doi.org/10.4039/n07-056>
- Hausmann A, Haszprunar G, Hebert PDN (2011) DNA barcoding the geometrid fauna of Bavaria (Lepidoptera): successes, surprises, and questions. *Public Library of Science ONE* 6: e17134. <https://doi.org/10.1371/journal.pone.0017134>
- Havemann N, Gossner MM, Hendrich L, Morinière J, Niedringhaus R, Schäfer P, Raupach MJ (2018) From water striders to water bugs: The molecular diversity of aquatic Heteroptera (Gerromorpha, Nepomorpha) of Germany based on DNA barcodes. *PeerJ* 6: e4577. <https://doi.org/10.7717/peerj.4577>
- Hawltischek O, Morinière J, Lehmann GUC, Lehmann AW, Kropf M, Dunz A, Glaw F, Detchaeron M, Schmidt S, Hausmann A, Szucsich NU, Caetano-Wyler SA, Haszprunar G (2017) DNA barcoding of crickets, katydids and grasshoppers (Orthoptera) from Central Europe with focus on Austria, Germany and Switzerland. *Molecular Ecology Resources* 17: 1037–1053. <https://doi.org/10.1111/1755-0998.12638>
- Hazakani-Covo E, Zeller RM, Martin W (2010) Molecular poltergeists: mitochondrial DNA copies (numts) in sequenced nuclear genomes. *Public Library of Science Genetics* 6: e1000834. <https://doi.org/10.1371/journal.pgen.1000834>



- Hebert PDN, Cywinska A, Ball SL, deWaard JR (2003a) Biological identifications through DNA barcodes. *Proceedings of the Royal Society of London Series B: Biological Sciences* 270: 313–321. <https://doi.org/10.1098/rspb.2002.2218>
- Hebert PDN, Ratnasingham S, deWaard JR (2003b) Barcoding animal life: cytochrome *c* oxidase subunit 1 divergences among closely related species. *Proceedings of the Royal Society of London Series B: Biological Sciences* 270: S96–S99. <https://doi.org/10.1098/rsbl.2003.0025>
- Hebert PDN, Gregory TR (2005) The promise of DNA barcoding for taxonomy. *Systematic Biology* 54: 852–859. <https://doi.org/10.1080/10635150500354886>
- Hendrich L, Morinière J, Haszprunar G, Hebert PDN, Hausmann A, Köhler F, Balke M (2015) A comprehensive DNA barcode database for Central European beetles with a focus on Germany: Adding more than 3,500 identified species to BOLD. *Molecular Ecology Resources* 15: 795–818. <https://doi.org/10.1111/1755-0998.12354>
- Hůrka K (1996) *Carabidae of the Czech and Slovak Republics*. Kabouek, Zlín, 565 pp.
- Hurst GDD, Jiggins FM (2005) Problems with mitochondrial DNA as marker in population, phylogeographic and phylogenetic studies: the effects of inherited symbionts. *Proceedings of the Royal Society Series B: Biological Sciences* 272: 1525–1534. <https://doi.org/10.1098/rspb.2005.3056>
- Ivanova NV, deWaard JR, Hebert PDN (2006) An inexpensive, automation-friendly protocol for recovering high-quality DNA. *Molecular Ecology Notes* 6: 998–1002. <https://doi.org/10.1111/j.1471-8286.2006.01428.x>
- Kajtoch Ł, Kolasa M, Kubisz D, Gutowski JM, Ścibior R, Mazur MA, Holecová M (2019) Using host species traits to understand the *Wolbachia* infection distribution across terrestrial beetles. *Scientific Reports* 9: e847. <https://doi.org/10.1038/s41598-018-38155-5>
- Kimura M (1980) A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution* 16: 111–120. <https://doi.org/10.1007/BF01731581>
- Kolasa M, Kubisz D, Mazur MA, Ścibior R, Kajtoch Ł (2018) *Wolbachia* prevalence and diversity in selected riverine predatory beetles (Bembidiini and Paederini). *Bulletin of Insectology* 71: 193–200.
- Krees WJ, Garcia-Robledo C, Uriarte M, Erickson DI (2015) DNA barcodes for ecology, evolution, and conservation. *Trends in Ecology and Evolution* 30: 25–35. <https://doi.org/10.1016/j.tree.2014.10.008>
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K (2018) MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Molecular Biology and Evolution* 35: 1547–1549. <https://doi.org/10.1093/molbev/msy096>
- Liebherr JK, Schmidt J (2004) Phylogeny and biogeography of the Laurasian genus *Agonum* Bonelli (Coleoptera, Carabidae, Platynini). *Deutsche Entomologische Zeitschrift* 51: 151–206. <https://doi.org/10.1002/mmnd.4810510202>
- Liebherr JK, Schmidt J, Lorenz W (2005) Nomenclatural correction to Liebherr and Schmidt's "Phylogeny and biogeography of the Laurasian genus *Agonum* Bonelli (Coleoptera, Carabidae, Platynini)." [see *Dtsch. Entomol. Z.* 51 (2004) 2, 151–206]. *Deutsche Entomologische Zeitschrift* 52(2): 291–291. <https://doi.org/10.1002/mmnd.4810520211>
- Leigh JW, Bryant D (2015) POPART: Full-feature software for haplotype network construction. *Methods in Ecology and Evolution* 6: 1110–1116. <https://doi.org/10.1111/2041-210X.12410>
- Lindroth CH (1986) The Carabidae (Coleoptera) of Fennoscandia and Denmark. *Fauna Entomologica Scandinavica* 15, part 2: 233–497.
- Luff ML (2007) The Carabidae (ground beetles) of Britain and Ireland (2<sup>nd</sup> edition). *Handbooks for the Identification of British Insects Vol. 4 Part 2*. Field Studies Council, Shrewsbury, 247 pp.
- Maddison DR (2012) Phylogeny of *Bembidion* and related ground beetles (Coleoptera: Carabidae: Trechinae: Bembidiini: Bembidiina). *Molecular Phylogenetics and Evolution* 63: 533–576. <https://doi.org/10.1016/j.ympev.2012.01.015>
- Miller SE, Hausmann A, Hallwachs W, Janzen DH (2016) Advancing taxonomy and bioinventories with DNA barcodes. *Philosophical Transactions of the Royal Society of London B: Biological Sciences* 371(1702): e20150339. <https://doi.org/10.1098/rstb.2015.0339>
- Morinière J, Hendrich L, Balke M, Beermann AJ, König T, Hess M, Koch S, Müller R, Leese F, Hebert PDN, Hausmann A, Schubart CD, Haszprunar G (2017) A DNA barcode library for Germany's mayflies, stoneflies and caddisflies (Ephemeroptera, Plecoptera and Trichoptera). *Molecular Ecology Resources* 17: 1293–1307. <https://doi.org/10.1111/1755-0998.12683>
- Mutanen M, Kivelä SM, Vos RA, Doorenweerd C, Ratnasingham S, Hausmann A, Huemer P, Dinca V, van Nieuwerkerken EJ, Lopez-Vaamonde C, Vila R, Aarvik L, Decaens T, Efetov KA, Hebert PDN, Johnsen A, Karsholt O, Pentinsaari M, Rougerie R, Segerer A, Tarmann G, Zahiri R, Godfray HCJ (2016) Species-level para- and polyphyly in DNA barcode gene trees: strong operational bias in European Lepidoptera. *Systematic Biology* 65: 1024–1040. <https://doi.org/10.1093/sysbio/syw044>
- Pail W (2010) *Agonum scitulum* Dejean, 1828 in Österreich – bisher übersehen oder in Ausbreitung begriffen? (Coleoptera: Carabidae). *Beiträge zur Entomofaunistik* 11: 79–83. [In German]
- Pentinsaari M, Hebert PDN, Mutanen M (2014) Barcoding beetles: a regional survey of 1872 species reveals high identification success and unusually deep interspecific divergences. *Public Library of Science ONE* 9: e108651. <https://doi.org/10.1371/journal.pone.0108651>
- Ratnasingham S, Hebert PDN (2007) BOLD: The Barcode of Life Data Systems. *Molecular Ecology Notes* 7: 355–364. <https://doi.org/10.1111/j.1471-8286.2007.01678.x>
- Ratnasingham S, Hebert PDN (2013) A DNA-based registry for all animal species: the Barcode Index Number (BIN) system. *Public Library of Science ONE* 8: e66213. <https://doi.org/10.1371/journal.pone.0066213>
- Raupach MJ, Astrin JJ, Hannig K, Peters MK, Stoeckle MY, Wägele JW (2010) Molecular species identifications of Central European ground beetles (Coleoptera: Carabidae) using nuclear rDNA expansion segments and DNA barcodes. *Frontiers in Zoology* 7: e26. <https://doi.org/10.1186/1742-9994-7-26>
- Raupach MJ, Hannig K, Morinière J, Hendrich L (2016) A DNA barcode library for ground beetles (Insecta: Coleoptera: Carabidae) of Germany: The genus *Bembidion* Latreille, 1802 and allied taxa. *ZooKeys* 592: 121–141. <https://doi.org/10.3897/zookeys.592.8316>
- Raupach MJ, Hannig K, Morinière J, Hendrich L (2018) A DNA barcode library for ground beetles of Germany: The genus *Amara* Bonelli, 1810 (Insecta: Coleoptera: Carabidae). *ZooKeys* 759: 57–80. <https://doi.org/10.3897/zookeys.759.24129>
- Raupach MJ, Hannig K, Morinière J, Hendrich L (2019) About *Notiophilus* Duméril, 1806 (Coleoptera, Carabidae): Species delin-



- eation and phylogeny using DNA barcodes. *Deutsche Entomologische Zeitschrift* 66: 63–73. <https://doi.org/10.3897/dez.66.34711>
- Raupach MJ, Hannig K, Wägele JW (2011) Identification of Central European ground beetles of the genus *Bembidion* (Coleoptera: Carabidae) using DNA barcodes: a case study of selected species. *Angewandte Carabidologie* 9: 63–72.
- Raupach M, Hendrich L, Küchler SM, Deister F, Morinière J, Gossner MM (2014) Building-up of a DNA barcode library for true bugs (Insecta: Hemiptera: Heteroptera) of Germany reveals taxonomic uncertainties and surprises. *Public Library of Science ONE* 9: e106940. <https://doi.org/10.1371/journal.pone.0106940>
- Ross HA (2014) The incidence of species-level paraphyly in animals: a re-assessment. *Molecular Phylogenetics and Evolution* 76: 10–17. <https://doi.org/10.1016/j.ympev.2014.02.021>
- Rubioff D, Cameron S, Will K (2006) A genomic perspective on the shortcomings of mitochondrial DNA for “barcoding” identification. *Journal of Heredity* 97: 581–594. <https://doi.org/10.1093/jhered/esl036>
- Rulik B, Eberle J, von der Mark L, Thormann J, Jung M, Köhler F, Apfel W, Weigel A, Kopetz A, Köhler J, Fritzlar F, Hartmann M, Hadulla K, Schmidt J, Hörren T, Krebs D, Theves F, Eulitz U, Skale A, Rohwedder D, Kleeberg A, Astrin JJ, Geiger MF, Wägele JW, Grobe P, Ahrens D (2017) Using taxonomic consistency with semi-automated data pre-processing for high-quality DNA barcodes. *Methods in Ecology and Evolution* 8: 1878–1887. <https://doi.org/10.1111/2041-210X.12824>
- Saitou N, Nei M (1987) The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* 4: 406–425.
- Schmid-Egger C, Straka J, Ljubomirov T, Blagoev GA, Morinière J, Schmidt S (2019) DNA barcodes identify 99 per cent of apoid wasp species (Hymenoptera: Ampulicidae, Crabronidae, Sphecidae) from the Western Palearctic. *Molecular Ecology Resources* 19: 476–484. <https://doi.org/10.1111/1755-0998.12963>
- Schmidt J (1994) Revision der mit *Agonum* (s. str.) *viduum* (Panzer, 1797) verwandten Arten (Coleoptera, Carabidae). *Beiträge zur Entomologie* 44: 3–51. [In German]
- Schmidt J (2006) 68. Gattung *Agonum* Bonelli, 1810. In: Freude H, Harde KW, Lohse GA, Klausnitzer B (Eds) *Die Käfer Mitteleuropas. Band 2, Adephaga I, Carabidae (Laufkäfer)*. Spektrum Akademischer Verlag, München, 262–279. [In German]
- Schmidt J, Benedikt S (2010) Zur Verbreitung des *Agonum scitulum* Dejean, 1828 (Coleoptera, Carabidae). *Entomologische Nachrichten und Berichte* 54: 64–65. [In German]
- Schmidt J, Trautner J, Müller-Motzfeld G (2016) Rote Liste und Gesamtartenliste der Laufkäfer (Coleoptera: Carabidae) Deutschlands. In: Gruttker H, Balzer S, Binot-Hafke M, Haupt H, Hofbauer N, Ludwig G, Matzke-Hajek G, Ries M (Eds) *Rote Liste der gefährdeten Tiere, Pflanzen und Pilze Deutschlands. Band 4: Wirbellose Tiere (Teil 2)*. Naturschutz und Biologische Vielfalt 70: 139–204. [In German]
- Schmidt S, Schmid-Egger C, Morinière J, Haszprunar G, Hebert PDN (2015) DNA barcoding largely supports 250 years of classical taxonomy: identifications for Central European bees (Hymenoptera, Apoidea *partim*). *Molecular Ecology Resources* 15: 985–1000. <https://doi.org/10.1111/1755-0998.12363>
- Schoville SD, Roderick GK, Kavanaugh DH (2012) Testing the “Pleistocene species pump” in alpine habitats: lineage diversification of flightless ground beetles (Coleoptera: Carabidae: *Nebria*) in relation to altitudinal zonation. *Biological Journal of the Linnean Society* 107: 95–111. <https://doi.org/10.1111/j.1095-8312.2012.01911.x>
- Sota T, Takami Y, Monteith GB, Moore BP (2005) Phylogeny and character evolution of endemic Australian carabid beetles of the genus *Pamborus* based on mitochondrial and nuclear gene sequences. *Molecular Phylogenetics and Evolution* 36: 391–404. <https://doi.org/10.1016/j.ympev.2005.02.015>
- Sota T, Vogler AP (2002) Incongruence of mitochondrial and nuclear gene trees in the carabid beetles *Ohomopterus*. *Systematic Biology* 50: 39–59. <https://doi.org/10.1093/sysbio/50.1.39>
- Takami Y, Suzuki H (2005) Morphological, genetic and behavioral analyses of a hybrid zone between the ground beetles *Carabus lewisianus* and *C. albrechti* (Coleoptera, Carabidae): asymmetrical introgression caused by movement of the zone? *Biological Journal of the Linnean Society* 86: 79–94. <https://doi.org/10.1111/j.1095-8312.2005.00527.x>
- Trautner J, Fritze M-A, Hannig K, Kaiser M (2014) *Distribution Atlas of Ground Beetles in Germany*. Books on Demand, Norderstedt, 348 pp.
- Weigand H, Beermann AJ, Čiampor F, Costa FO, Csabai Z, Duarte S, Geiger MF, Grabowski M, Rimet F, Rulik B, Strand M, Szucsich N, Weigand AM, Willassen E, Wyler SA, Bouchez A, Borja A, Čiamporová-Zatovičová Z, Ferreira S, Dijkstra S-DB, Eisendle U, Freyhof J, Gdawski P, Graf W, Haegerbaeumer A, van der Hoorn BB, Japoshvili B, Keresztes L, Keskin E, Leese F, Macher JN, Mamos T, Paz G, Pešić V, Pfannkuchen DM, Pfannkuchen MA, Price BW, Rinkevich B, Teixeira MAL, Várbiro G, Ekrem T (2019) DNA barcode reference libraries for the monitoring of aquatic biota in Europe: Gap-analysis and recommendations for future work. *Science of the Total Environment* 678: 499–524. <https://doi.org/10.1016/j.scitotenv.2019.04.247>
- Weng Y-M, Veire BM, Dudko RY, Medeiros MJ, Kavanaugh DH, Schoville SD (2020) Rapid speciation and ecological divergence into North American alpine habitats: the *Nipponebria* (Coleoptera: Carabidae) species complex. *Biological Journal of the Linnean Society* 130: 18–33. <https://doi.org/10.1093/biolinnean/blaa014>
- Weng Y-M, Yang M-M, Ye W-B (2016) A comparative phylogeographic study reveals discordant evolutionary histories of alpine ground beetles (Coleoptera, Carabidae). *Ecology and Evolution* 6: 2061–2073. <https://doi.org/10.1002/ece3.2006>
- Will KW, Mishler BD, Wheeler QD (2015) The perils of DNA barcoding and the need for integrative taxonomy. *Systematic Biology* 54: 844–851. <https://doi.org/10.1080/10635150500354878>
- Will KW, Rubioff D (2004) Myth of the molecule: DNA barcodes for species cannot replace morphology for identification and classification. *Cladistics* 20: 47–55. <https://doi.org/10.1111/j.1096-0031.2003.00008.x>
- Zhang AB, Kubota K, Takami Y, Kim JL, Kim JK, Sota T (2005) Species status and phylogeography of two closely related *Coptolabrus* species (Coleoptera: Carabidae) in South Korea inferred from mitochondrial and nuclear gene sequences. *Molecular Ecology* 14: 3823–3841. <https://doi.org/10.1111/j.1365-294X.2005.02705.x>



## Supplementary material 1

### Barcode analysis using the BOLD workbench

Authors: Michael J. Raupach, Karsten Hannig, Jérôme Morinière, Lars Hendrich

Data type: Data table

Explanation note: Molecular distances are based on the Kimura 2-parameter model of the analysed specimens of the analysed species of the genus *Agonum* and *Oxytelus obscurus* (Paykull, 1790). Divergence values were calculated for all studied sequences, using the Nearest Neighbour Summary implemented in the Barcode Gap Analysis tool provided by the Barcode of Life Data System (BOLD). Align sequencing option: BOLD aligner (amino acid-based HMM), ambiguous base/gap handling: pairwise deletion. ISD = intraspecific distance. BINs are based on the barcode analysis from 16-04-2020. Species pairs with interspecific distances < 2.2% are marked in bold. As a consequence of that the number of unspecified nucleotides (“Ns”) exceeds more than 1% of the total length of the sequence, barcodes of *Agonum antennarium* (Duftschmid, 1812) and *A. ericeti* (Panzer, 1809) received no BIN. Country codes: D = Germany, A = Austria, B = Belgium, BG = Bulgaria, CZ = Czech Republic, EST = Estonia, FIN = Finland, F = France, I = Italy, RO = Romania, SK = Slovakia, SLO = Slovenia, and SW = Switzerland.

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## Supplementary material 2

### Neighbour-joining topology

Authors: Michael J. Raupach, Karsten Hannig, Jérôme Morinière, Lars Hendrich

Data type: Neighbour-joining topology

Explanation note: Neighbour-joining topology of all analysed carabid beetles based on Kimura 2-parameter distances. Specimens are classified using ID numbers from BOLD and species name. Numbers next to nodes represent non-parametric bootstrap values (1,000 replicates, in %). Three beetles of a previous publication that were identified as *Agonum ericeti* (Panzer, 1809) (see Pentinsaari et al. 2014) were incorrectly determined. A careful re-inspection revealed these specimens as *Agonum sexpunctatum* (Linné, 1758).

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